

猕猴桃研究进展(VI)

Advances in Actinidia Research (VI)

黄宏文 主编Edited by Huang Hongwen

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内容简介

本书收录了国内猕猴桃会议代表提交的论文,以及近两年国外猕猴桃研究的新动态论文,内容涉及从产业、栽培管理技术、病虫害防治、生物技术采后贮藏加工、遗传育种到资源利用的7个主要领域。所录论文是国内外近年来从事猕猴桃研究、管理、开发利用人员的成果或工作积累,以及针对一些产业发展问题和新技术应用提供建议。

本书是供广大从事猕猴桃科研、教学、推广与生产、市场销售等领域人员参考的重要资料,适合科研人员、教师、大中专学生、职业院校及从事果树行业管理的行政部门人员、基层科技人员,以及猕猴桃爱好者阅读。

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前 言

(PREFACE)

猕猴桃(Actinidia chinensis Planch.)是 20 世纪初以来驯化栽培成功的水果,至今仅有 100 余年的历史。猕猴桃以其独特的风味,富含维生素 C、饮食纤维、多种矿物营养,以及清肠健胃功效而得到广泛青睐,成为重要的水果种类之一。猕猴桃的驯化栽培被认为是近代由野生到人工商品化栽培最成功的植物驯化范例。

我国自1978年开展全国猕猴桃属植物野生资源普查以来,经过30余年的发展,充分利用自己的资源优势大步追赶世界先进水平,现已成为世界栽培面积第一、产量第二的猕猴桃生产大国。至2010年,我国栽培面积达7.0万hm²,占世界栽培总面积的45%;产量达49.0万t,占世界产量的28%。我国猕猴桃产业的快速发展进程凝结了我国猕猴桃科技人员、果农及经销者多年的拼搏和不懈的追求!也正是为了这个追求,中国园艺学会猕猴桃分会于2004年开始,每2年召开1次全国学术与产业研讨会,已分别在湖南吉首、广东和平、陕西杨陵和四川蒲江举办了四届,每次会议之后均出版了《猕猴桃研究进展》系列学术论文集,本卷是继《猕猴桃研究进展(V)》之后的第VI卷,是在收集第四届猕猴桃研讨会会议论文的基础上,增加了世界主要生产国的知名专家在猕猴桃上的科研和产业等方面的研究论文,旨在为中外研究人员提供世界猕猴桃研究进展和市场、生产最新信息。

本书共分新品种选育、资源开发与利用、栽培及生理、生物技术、贮藏保鲜与加工、产业与市场6个部分,系统地提供了国内外猕猴桃科研与产业、市场发展趋势,也结合我国猕猴桃发展现状和存在问题,提出了我国猕猴桃科研重点、产业方向和市场策略。

由于能力和水平所限,书中疏漏之处在所难免,恳请大家批评指正。借此机会再次向为本书提供文献的作者和对分会和本书给予支持的领导和同仁们表示衷心的感谢,并希望继续得到你们更多的指导和支持!

中国园艺学会猕猴桃分会2011.6.10于武汉

目 录

(CONTENTS)

前言(PREFACE)

(一)新品种选育

| 利用多因子分析鉴定二倍体中华猕猴桃品种果实品质相关挥发性物质种类(Identifying |
|--|
| Volatile Compounds Associated with Sensory and Fruit Attributes in Diploid Actinidia chinensis |
| (kiwifruit)Using Mutivariate Analysis) |
| 猕猴桃育种中的倍性操作—2 倍体中华猕猴桃品种的染色体离体加倍技术(Manipulation of |
| Ploidy for Kiwifruit Breeding: in Vitro Chromosome Doubling in Diploid Actinidia chinensis |
| Planch.) |
| 极耐贮、晚熟黄肉猕猴桃新品种'金艳'('Jinyan': A Superior Yellow-fleshed Kiwifruit Cultivar |
| with Excellent Storage Quality) |
| 彩色猕猴桃两性花品种的研究(Research on Hermaphrodite Flower Variety of Actinidia deliciosa |
| var. coloris) 王明忠 等 41 |
| 中华无籽猕猴桃新品种'湘吉红'选育与栽培技术研究初报(Breeding and Ecological Cultivation |
| Techniques of a New Strain 'Xiangji-Hong' of Seedless Actinidia chinensis) 裴昌俊 等 46 |
| 全红型软枣猕猴桃品种'天源红'的选育(New All-red Kiwifruit Cultivars: 'Tianyuanhong') |
| |
| 适合江苏生态区域种植的红心猕猴桃杂交新品种选育(Cultivar Development of Red-flesh |
| Kiwifruit Forecological Environment Planting in Jiangsu Province) · · · · · 杨声谋 等 51 |
| 美味猕猴桃特早熟品种'海艳'的选育报告(Selection of a New Early-ripe Kiwifruit Cultivar- |
| 'Haian', from Actinidia chinensis var. deliciosa) ···································· |
| 中华猕猴桃黄肉新品种'豫皇 1 号'选育研究(Selection of a New Yellow-fleshed Cultivar |
| 'Yuhuang No.1', from Actinidia chinensis var. chinensis) ······ 田志刚 等 58 |
| 中华猕猴桃金黄果肉新品种'豫皇 2 号'选育研究(Selection of a New Golden-fleshed Cultivar |
| 'Yuhuang No. 2', from Actinidia chinensis var. chinensis) ····· 田志刚 等 62 |
| 晚熟中华猕猴桃新品种'皖金'的选育(A New Late Maturing Kiwifruit Cultivar:'Wanjin') |
| |
| 美味无籽猕猴桃新品种'湘吉'的选育与丰产栽培技术研究(Selection and Cultivation Technique |
| of a New Seedless Cultivar 'Xiangji'from Actinidia deliciosa) 裴昌俊 等 69 |
| (二)资源开发利用 |
| 猕猴桃植物新品种保护及转让现状(Study on the Protection and Transfer of New Kiwifruit |
| |

Varieties) 陈庆红 等 75

| 珍稀濒危植物金花猕猴桃优势群落结构特征分析(Analysis on the Stucture Charscteristcs of the |
|---|
| Dominant Community of Actinidia chrysantha) ······ 龚弘娟 等 80 |
| 红肉猕猴桃品种开发与市场潜力探讨(Red-fleshed Kiwifruit Cultivar Development and Its |
| Commercial Potential in China) ······ 姜正旺 等 86 |
| 辽宁地区软枣猕猴桃果实性状、次生物质鉴定及营养分析(The Actinidia arguta Fruit Character, |
| Secondary Substance Identify and Nutrient Analyses in Liaoning) 刘延吉 等 91 |
| 东北野生猕猴桃资源普查保护与开发利用(Studies on the Wild Actinidia arguta Resources |
| Protection and Exploitation of Investigation in Northeast China) ···································· |
| 软枣猕猴桃种质资源遗传多样性研究(Study on Genetic Diversity of Actinidia arguta Germplasm |
| Resources) |
| |
| (三)栽培及生理 |
| 基于 16S-23S 核糖体 DNA 内转录间隔区间(ITS)和其他基因区间 PCR 引物比对,获得 PCR 引 |
| 物用以鉴别 PSA(猕猴桃溃疡病)不同菌株(Detection of Pseudomonas syringae pv. actinidiae |
| Using Polymerase Chain Reaction (PCR) Primers Based on the 16S-23S rDNA Intertranscribed |
| Spacer Region and Comparison with PCR Primers Based on other Gene Regions) |
| |
| 猕猴桃功能性花粉发育中多胺类物质的生物合成与调控(Polyamine Biosynthesis and Control of |
| the Development of Functional Pollen in Kiwifruit) ····· G. Falasca 等 125 |
| 采前除袋对'金艳'猕猴桃成熟度及贮藏期品质变化的影响(Effects of Pre-harvest Debagging on |
| Maturity and Quality Change in Storage of 'Jinyan' Kiwifruit) 陈美艳 等 140 |
| '金魁'猕猴桃适配雄株筛选试验研究(A Study on Selection of Superior Male Plant for Actinidia |
| chinensis var. deliciosa 'Jinkui') ····· 顾 霞等 143 |
| 猕猴桃不同雄性品系染色体倍性与花性特征相关性研究(Study on Blooming and Pollen |
| Germination in Kiwifruit Pollenizers of Different Ploidy Levels) · · · · · · 韩 飞 等 148 |
| 不同雄株花粉对'华优'果实的影响(Effect of Male Vines to the Fruiting of 'Huayou' Kiwifruit) |
| |
| 猕猴桃新害虫:斜纹夜蛾的研究(Study of a New Kiwifruit Pest Prodenia litura) |
| |
| 渍水处理对美味猕猴桃生理生化特性影响(Physiological and Biochemical Responses to Water |
| Logging in Actinidia deliciosa cv Jinkui) ······ 孙丹柯 等 158 |
| '华优'猕猴桃栽培技术(Cultivation Techniques for 'Huayou' Kiwifruit) 王西锐 等 162 |
| 猕猴桃溃疡病防治研究(Study on the Control Techniques for Kiwifruit Bacterial Canker) |
| |
| 猕猴桃细菌性溃疡病综合防治技术(The Integrated Control Technique for Kiwifruit Bacterial |
| Canker) ····· 王西锐 等 171 |

| 不同营养液浓度对'徐香'猕猴桃果实发育和主要营养物质含量的影响(Effects of Nutrient |
|--|
| Solution of Different Concentrations on Development and Mail Nutriment Contents 'Xuxiang' |
| Kiwifruit) ····· 蔚玉红 等 174 |
| 猕猴桃叶片矿质元素变化规律及其与果实品质关系的研究(Study on the Change Pattern of |
| Mineral Elements in Kiwifruit Leaves and Relation with Fruit Quality) 徐爱春 等 179 |
| 红阳猕猴桃果实套袋试验(Fruit Bagging Test of 'Hongyang') ····· 余中树 等 187 |
| '红阳'猕猴桃不良表现及应对措施(The Problems and Solutions of 'Hongyang') |
| 余中树 189 |
| 冰冻灾害对庐山植物园猕猴桃生长发育与产量的影响(The Effects of Cold Injury on the Growth |
| and Yield of Actinidia in Lushan Botanical Garden) ······ 虞志军等 193 |
| 猕猴桃品种'金桃'和'金艳'果实发育规律研究(Study on the Regulation of Fruit Development in |
| Kiwifruit Cultivars-'Jintao' and 'Jinyan') ···································· |
| |
| (四)生物技术 |
| 意大利中部获得的黄肉猕猴桃品种近期流行的猕猴桃溃疡病菌的分子及形态特征(Molecular |
| and Phenotypic Features of <i>Pseudomonas syringae</i> pv. actinidiae Isolated During Recent Epidemics |
| of Bacterial Canker on Yellow Kiwifruit (Actinidia chinensis) in Central Italy) |
| |
| 猕猴桃种间杂交后代不同倍性个体 Vc,花色和倍性变异及 SSR 分析(Vitamin C, Flower Color |
| and Ploidy Variation of Hybrids from a Ploidy-unbalanced Actinidia Interspecific Cross and SSR |
| Characterization) ···································· |
| 猕猴桃等果树抗坏血酸研究进展(Advances in Research of Ascorbic Acid on Kiwifruit and other |
| Fruit Trees) ····· 高 洁 等 229 |
| 海拔和经度连续渐变区域中华猕猴桃复合体形态和倍性变异研究(Morphological and Cytotype |
| Variation of Wild Kiwifruit (Actinidia chinensis complex) Along an Altitudinal and Longitudinal |
| Gradient in Central-west China) ···································· |
| 秦岭区域中华/美味猕猴桃复合体遗传多样性及其景观遗传结构(Genetic Diversity and |
| Landscape Genetic Structure of Actinidia chinensis var. chinensis/deliciosa Complex in the Qinling |
| Mountains) · · · · · 廖 利 等 251 |
| 辽宁地区5种野生软枣猕猴桃 RAPD 遗传多样性分析(A Primary Analysis on Random Amplified |
| Polymorphic DNA in Five Actinidia arguta Species from Liaoning Province) ··· 刘延吉 等 270 |
| 不同栽培环境下'红阳'内果皮着色的细胞学差异(Cytological Mechanism of Coloration |
| Differences in Inner Pericarp of 'Hongyang' Cultivated in Different Conditions) |
| |
| 基于 NCBI 公共数据平台的软枣猕猴桃 EST-SSR 开发(Exploitation of EST-SSRs in Actinidia |
| |

| 毛花猕猴桃果实 L-半乳糖脱氢酶(GalDH)cDNA 的克隆与序列分析(Cloning and Sequence |
|--|
| Analysis of cDNA of L-galactose Dehydrogenase from Fruit of Actinidia eriantha) |
| |
| 基于猕猴桃 EST 序列的 SSR 引物开发(Mining of SSR Primers Based on EST Sequences in |
| Kiwifruit)徐小彪 等 289 |
| ⁶⁰ Co-γ 对三种猕猴桃辐射诱变适宜剂量的研究(Search for Proper Dose of ⁶⁰ Co-γ Ray in Three |
| Varieties of Kiwifruit Radiation Breeding) ···································· |
| SSR 标记在猕猴桃属两个 F1群体中的分离研究(Inheritance of SSR Markers in Two F1 Hybrid |
| Population of Actinidia) ···································· |

(五)贮藏保鲜与加工

贮藏猕猴桃果肉硬度测试中穿刺速度对测定结果的影响(Effect of Penetration Speed on Flesh 新型商业开发软枣猕猴桃及其杂种果实采后性状与不同基因型间的差异分析 I. 品质鉴评特 性与理化差异(Genotypic Difference in Postharvest Characteristics of Hardy Kiwifruit(Actinidia arguta and its Hybrids), as a New Commercial Crop Part I. Sensory Profiling and Physicochemical Differences) Piotr Latocha 等 324 '金艳'猕猴桃后熟过程中的品质变化(Quality Changes of'Jinyan'Kiwifruit with Different Harvest Maturity during their Afterripening) · · · · 李昆同 等 341 浅析川渝'红阳'猕猴桃采收、运输中存在的问题(A Brief Analysis of Problems Existed in Harvesting and Transportation of 'Hongyang' Kiwifruit in Sichuan & Chongqing) 李昆同 等 346 软枣猕猴桃多糖降血糖降血脂活性研究(Polysaccharide of Actinidia arguta and Activity of Blood 软枣猕猴桃多糖提取工艺及分离纯化研究(Study on the Extraction,Separation and Purification of 采收期和贮藏温度对'金艳'猕猴桃品质的影响(Effects of Harvest Date and Storage Temperature on Quality of 'Jinyan' Kiwifruit) ······ 钱政江 等 361 猕猴桃果实耐贮性的主要影响因子研究 (Main Effects of Storage Characteristic of Kiwifruit Fruits) ······ 王仁才 等 369

(六)产业与市场

决定消费者购买意向:猕猴桃果实干物质含量、大小和价格(Determining Consumer Purchase Intentions: the Importance of Dry Matter, Size, and Price of Kiwifruit) ··· Sara R. Jaeger 等 377 当前影响我国猕猴桃产业健康发展的主要因素和对策(The Main Factors Influencing the Current Kiwifruit Industry and Strategies for Sustainable Development in China) ··· 對仁煌 等 391 · vi ·

(一)新品种选育

利用多因子分析鉴定二倍体中华猕猴桃品种果实品质 相关挥发性物质种类

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摘 要 在对果实中的糖、酸类物质影响风味进行了很多关注的同时,往往对果实风味与香气同样有重要作用的挥发性物质却难以定义。那些对中华猕猴桃品种风味有重要贡献挥发性物质的鉴定,可以大大提高果实风味改良育种的效率与降低成本。本研究就是利用多因子分析来探讨挥发性化合物、品质鉴评和中华猕猴桃品种果实特性来确定影响风味的主要挥发性物质,以及优良亲本及选择方法。我们从一个育种群体中利用主成分分析(PCA)方法,依据果实特性和风味多样性相关性状为原则选择了24个基因型进行测试,发现了72个挥发性物质。事实证明,利用扩展的多因子分析可以从有限的材料中发掘最大量的信息,在主成分分析中,为避免其中单因子相关矩阵分析的缺陷,我们基于挥发性物质表型相关多因子进行聚类分析,成功将这些物质被聚类成4大家族。根据这些分支,进一步利用 PCA 和多因子线性回归分析,探讨了挥发性物质、感官评价和果实性状间的相关性。主成分分析提供了对影响消费者反应的挥发性物质进行综合权衡的依据,13种对中华猕猴桃品种果实品质有重要影响的挥发性物质被鉴定,'Hort16A'中有5种脂类对其果实风味和香气有重要决定性作用。与"酸味"、"果实成熟"、"Hort16A 非典型性香气"和"非猕猴桃味(一般指'海沃德')"相关的挥发性物质也鉴定出来。同时,对具备不同挥发性物质的潜在亲本选择以及猕猴桃的风味改良选育的可行性方法也进行了总结。

关键词 猕猴桃 育种 风味 挥发性物质 多因子分析

Identifying Volatile Compounds Associated with Sensory and Fruit Attributes in Diploid *Actinidia chinensis*(kiwifruit) Using Mutivariate Analysis

1 Introduction

Kiwifruit belong to the genus *Actinidia*, which comprises more than 60 species native to large parts of China and some neighbouring countries. All known species are dioecious and grow as long-lived, perennial, woody vines. The familiar green-fleshed kiwifruit of the cultivar 'Hayward' belongs to the species *A. deliciosa* (A. Chev.) Liang et A. R. Ferguson and is grown commercially in many countries. Yellow-fleshed cultivars of *A. chinensis* Planch. are grown widely in China and increasingly in other kiwifruit producing countries. The 'Hayward' cultivar is characteristically described by consumers as having a fresh, sweet and acid flavour, while the commercial yellow-fleshed *A. chinensis* cultivar 'Hort16A' (marketed as ZESPRI® GOLD Kiwifruit) is described as having sweet, banana and blackcurrant-like flavours (Jaeger et al., 2003).

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Flavour is a key aspect of fruit quality and is generally defined as a combination of aroma and taste sensations. While the roles of sugars and acids in taste are well known and described, the volatile compounds that contribute to flavour and odour are more difficult to define. Many of the earlier studies on kiwifruit aroma focused on the composition of volatile compounds. Up to 90 volatiles were identified from fruit of 'Hayward' (Young et al., 1983; Young and Paterson, 1985; Takeoka et al., 1986; Paterson et al., 1991; Young and Paterson, 1995; Young et al., 1995; Perera et al., 1998). Between 40 and 80 volatiles were detected from fruit of A. arguta genotypes (Matich et al., 2003). Hex-E2-enal was identified as the major component in mature fruit of 'Hayward' but on further ripening, ethyl butanoate began to dominate the profile (Bartley and Schwede, 1989). Young et al. (1995) reported that the sweetness of ripe 'Hayward' kiwifruit was associated with high concentrations of esters, which account for up to 85 % of the total volatiles in kiwifruit (Crowhurst et al., 2008). In the yellow-fleshed cultivar 'Hort16A', accetaldehyde, hexanal, hex-E2-enal and ethyl butanoate were found to be important contributors to aroma (Friel et al., 2007).

However, because many volatile compounds are not flavour active in a particular product, one of the major difficulties in studying odour is the determination of those compounds that make a real contribution to the flavour of food (Mistry et al., 1997). Odour activity values (OAVs) are used to estimate the odour contribution of individual food volatiles, and are calculated as the ratio between the concentration of an individual substance in a sample and the threshold concentration of that substance (i. e., the minimal concentration that can be detected by the human nose) (Rothe and Thomas, 1963). Buttery (1993) found that of 400 volatiles identified in tomatoes, only 16 reached their OAVs. Hundreds of volatiles have also been identified in apple (Dixon and Hewett, 2000) and strawberry (Dirinck et al., 1981; Pérez et al., 1996), but of these, only a small proportion was classified as 'character impact' volatiles. Of 60 volatiles identified from 'Hayward' kiwifruit using GC/MS-O(gas chromatography/mass spectrometryolfactometry), only 30 were odour-active (Frank et al., 2007). Hex-E2-enal ('marzipan, sweet') had the greatest odour impact, followed by 1-penten-3-one ('plastic, herbal, glue, kiwifruit-like') and hexanal ('green, fresh crushed leaves'). In 'Hayward' essence and puree, hex-E2-enal was perceived by GC-O as imparting a fruity, strawberry and cherry flavour (Jordán et al., 2002).

Olfactometry is thus a valuable tool as it allows a complex volatile mixture to be separated temporally into individual compounds, but it does not take account of the complexity of human sensory perception (Frank et al., 2007). Sensory evaluation methods offer an organized way to collect information on the sensory attributes of food samples as perceived by the human senses. To take into account the multivariate nature of the data, multivariate chemometric data analysis techniques ought to be applied (Daszykowski et al., 2007). Thus, a complementary approach to studying flavour is to use multivariate statistics to identify associations between sensory and chemical data that may indicate potentially important flavour volatiles that contribute most to the sensory experience. This approach recognizes that while individual flavour components are responsible for taste and odour, the total sensation determined from sensory analysis results from the integration of all the individual flavour stimuli in a mixture (Rouseff and Leahy, 1995; Frank et al., 2007). This type of approach has been used in studies of tomato, where relationships between sensory attributes and volatiles have been identified using multiple linear regression (MLR) (Baldwin et al., 1998; Tandon et al., 2003) and PCA(Krumbein et al., 2004).

Variation has been reported in the quantity of volatiles in thirteen apple cultivars (Young et al., 2004)

and six apricot cultivars (Guillot et al.,2006). López et al. (1998) found that odour components varied in class and quantity in two apple cultivars and were associated with sensory properties characteristic of each cultivar. During fruit ripening, a number of different biosynthetic pathways are involved in volatile synthesis (Dixon and Hewett,2000; González-Agüero et al.,2009; Zhang et al.,2009). These have not been fully described, but appear to be common to different fruits (Dixon and Hewett,2000). To date, only a few genes that directly influence fruit flavour biogenesis have been reported, including alcohol dehydrogenase (ADH2) in tomato (Speirs et al., 1998), terpene synthases in strawberry (Aharoni et al., 2004), antisense ACC (1-aminocyclopropane carboxylic acid) oxidase and alcohol acyltransferase (AAT) in apple (Schaffer et al., 2007; Souleyre et al., 2005; Li et al., 2006) and lipoxygenase (LOX) in kiwifruit (Zhang et al., 2009).

In an ever more competitive and global market, flavour quality is increasingly important to consumers. Therefore, knowledge of kiwifruit flavour is of utmost importance in developing new cultivars. While studies of the composition of volatile compounds have detected up to 90 volatiles in kiwifruit cultivars and genotypes, there are only a few published studies on 'flavour impact' volatiles related to sensory attributes in 'Hayward' (Frank et al., 2007; Jordán et al., 2002). Moreover, because of the large number of volatiles, the assessment of the volatiles contributing to flavour within breeding populations is especially challenging. Therefore, the main goal of this study was to evaluate multivariate analyses of associations between volatiles, sensory and fruit attributes to identify potential key impact volatiles that make a substantial contribution to the flavour and odour of *A. chinensis* fruit, using 24 genotypes representing the diversity of taste and fruit characteristics in a breeding population. The study also aimed to identify economic and efficient breeding methods for improving flavour volatiles and superior parents for use in our breeding programme.

2 Materials and methods

2.1 Plant material

Twenty-four genotypes of diploid A. chinensis were sampled in April 2001 and were numbered from P1 to P24. Twenty-two were seedlings chosen from among 289 females in a population derived from a factorial mating design (3 females × 13 males), planted in 1994 at the Te Puke Research Centre, New Zealand. The other two genotypes were female parents of this population, Female B (P23) and Female C (P24), respectively. The third female parent, Female A, was missing from our study. The three female parents (A, B and C) were selected as testers based on results from previous studies (Cheng et al., 2004). Both Female A and Female B had moderate dry matter (DM) and soluble solids content (SSC). Female C was 'Hort16A', a yellow-fleshed cultivar with large fruit renowned for their good flavour (Jaeger et al., 2003). The 13 male parents were selected randomly from our germplasm collection. The 22 seedling genotypes were selected to be broadly representative of the population, based on the first two principal components (PCs) derived from 14 variables analysed in a previous study (Cheng et al., 2004). The variables included four sugars (fructose, glucose, sucrose and myo-inositol), three organic acids (malic, quinic and citric), fruit pH, titratable acidity, vitamin C and four fruit characters (fruit number per vine, fruit weight, DM and SSC at eating ripeness). The two PCs were associated with the traits "sweet taste factor" and "sour taste factor", and accounted for 50.7 % of the total variance. The 22 genotypes selected showed extreme differences in combinations of the two PCs. This approach was expected to maximise the discrimination of phenotypes among vines in this population.

From 50 to 60 fruit were harvested at random from each vine when the mean SSC of a three-fruit sample reached 10 %. SSC and DM were measured at harvest as described in Cheng et al. (2004). The remaining fruit were cool stored at 1.5 % before sensory and chemical analyses were performed.

2.2 Fruit sampling and chemical analysis

After 1 month of storage, fruit were removed from the cool store and ripened at 20 °C for 3 ~ 5 days. Fifteen fruit (firmness $0.5 \sim 0.7 \text{ kgf}$) from each vine were selected for sensory analysis. Each end of each fruit was sampled for SSC, and a longitudinal slice (1/3) was removed for chemical analysis. One half of this slice was used for sugar (fructose, glucose, sucrose and *myo*-inositol) and acid (malic, quinic and citric acids) analysis and the other half was sampled for volatile analysis. The remaining part of the fruit was used for sensory analysis (see below). The fruit samples for sugar and acid analysis were frozen in liquid nitrogen immediately and stored at -20 °C prior to analysis. The individual sugar and acid contents were measured by GLC (gas-liquid chromatographic) analysis as described by Cheng et al. (2004).

For volatile analysis, the methods of Matich et al. (2003) were used with minor modifications. Headspace volatiles released from 1 to 1.5 g of the pulped tissue were collected with a Chromosorb 105 absorbent trap (100 mg per trap) for 15 min at 23 °C at a flow rate of 25 ml min ⁻¹. The sampled traps were stored at -15 °C and analysed using GC-MS(HP5890-VG70) with a DB-wax column (J&W, 30 m × 0.25 mm × 0.25 μ m), within 2 weeks. Peaks were identified and quantified as described by Matich et al. (2003). All analyses were performed on triplicate tissue samples.

2.3 Sensory analysis

A panel of eight in-house assessors, trained to evaluate the flavour of kiwifruit, was selected (Marsh et al.,2006; Wang et al.,2010). Sensory analysis was performed in a sensory panel room at 20 °C. Sensory attributes were recorded for sweet taste, sour taste, 'Hort16A'-like flavour and odour and ethyl butanoate flavour and odour. The attribute intensities were recorded on 150 mm unstructured line scales anchored at zero for absent and 150 for extreme, using CompusenseTM 5 sensory software. The attributes were calibrated using 'Hort16A' fruit for both 'Hort16A'-like flavour and odour(100), 20 g 1 ⁻¹ and 40 g 1 ⁻¹ sucrose solutions for sweet taste(40 and 100, respectively), and 1 g 1 ⁻¹ malic acid solution for sour taste (115) as in Marsh et al. (2006). Sensory samples of fruit were presented monadically to panellists stemend up in coded plastic crème cups. Each sample was cut into three slices. The stem end and middle slice were used for odour and flavour assessment, respectively. The remaining third was used for texture assessment (data not shown). The eight assessors analyzed 24 genotypes of kiwifruit during six sessions. The final sensory data were averaged across the eight assessors.

2.4 Statistical and multivariate analysis

In this study, there were 72 volatile compounds (attributes), but only 24 genotypes (observations). Thus, the dimensions of the data matrix were 24 × 72. For PCA, the correlation matrix for the data was singular and some scoring coefficients were zero (Daszykowski et al., 2007). To overcome this problem, the volatile data were regrouped, using hierarchical clustering of variables as implemented by PROC VARCLUS (SAS Institute Inc., 2003), based on a phenotypic correlation matrix between volatiles estimated by PROC CORR. The VARCLUS procedure divided volatile variables into hierarchical clusters. Four clusters were arbitrarily chosen, to make the number of volatile attributes within the cluster equal to or less than the number of observations (24) (Anderberg, 1973). The four clusters had 19, 13, 20 and 19 volatiles,

respectively(Table 1). In Cluster 1, because four ketones (2-methylpentan-3-one, 4-methyl-2-hexanone, 4-methylpentan-2-one and 5-methylhexan-2-one) and butyl acetate were found only in P2, some scoring coefficients of the PCA were zero because of the singular correlation matrix. Thus, only butyl acetate was included in Cluster 1 for PCA (Table 1). A PCA was carried out on the volatiles in each of the four clusters, using the PRINCOMP procedure of SAS(SAS Institute Inc., 2003). In accordance with Morrison (1990), the analysis was performed on standardised data. The un-rotated structure of each analysis was used to interpret the meaning of the components because the emphasis was on ordination rather than structure explanation. To minimize the number of principal components used for each character ('parsimony') (Morrison, 1990), we set the cut-off at 70 % of the total variance. This meant it was necessary to consider only 14 principal components (PCs) for all four clusters (Table 2).

Multiple linear regressions (MLR) of DM, sugars, acids and sensory attributes were conducted as a function of the 14 PCs, to explore associations between volatile compounds, sensory and fruit attributes and to identify potential key 'flavour impact' volatiles of fruit of A. chinensis. MLR was carried out using PROC REG with the STB (standardized β) option of SAS (SAS Institute Inc., 2003). A standardisation for the regression coefficient (STB) is the process whereby raw data are transformed into new variables with a mean of 0 and a standard deviation of 1. DM and SSC were very highly correlated and showed very similar correlations with other taste and fruit characters (Cheng et al. 2004). Therefore, this study reports only the relationship between DM and volatiles. The combined analysis of PCA and MLR enabled 13 potential key 'flavour impact' volatiles in A. chinensis fruit to be identified. Another PCA was carried out with only the 13 volatiles, to enable differentiation of genotypes. The parsimony criterion subsequently led to the first two PCs being used. MLRs of sensory attributes were conducted as a function of the two PCs of key volatiles, to specify the relationship between key volatiles and sensory attributes and to identify superior parents for flavour breeding.

Table 1 Occurrence and relative abundance of volatile compounds found in 24 genotypes of Actinidia chinensis

| C 1 | Occurrence | Mean | Minimum | Maximum | 'Hort16A' | Cl | |
|-----------------|-------------|--|---------|--------------------------|-------------------|-------------|--|
| Compound | (genotypes) | /(ng g $^{-1}$ FW) /(ng g $^{-1}$ l | | /(ng g ⁻¹ FW) | $/(ng g^{-1} FW)$ | Cluster no. | |
| Aldehyde | | | | | | | |
| Acetaldehyde | 15 | 6.37 | 0 | 42.27 | 5.28 | 4 | |
| Propanal | 24 | 3.97 | 0.83 | 7.83 | 3.60 | 1 | |
| Butanal | 22 | 0.99 | 0 | 2.60 | 0 | 2 | |
| 2-methylbutanal | 3 | 0.21 | 0 | 2.54 | 0 | 1 | |
| Hexanal | 24 | 128.95 | 3.94 | 616.98 | 28.24 | 4 | |
| Hex-E3-enal | 22 | 4.76 | 0 | 23.93 | 0 | 4 | |
| Hex-Z3-enal | 21 | 19.84 | 0 | 433.16 | 0 | 3 | |
| Heptanal | 23 | 2.93 | 0 | 11.42 | 1.82 | 1 | |
| Hex-Z2-enal | 22 | 11.74 | 0 | 39.01 | 4.45 | 4 | |
| Hex-E2-enal | 24 | 686.48 | 17.15 | 2 484.81 | 123.50 | 2 | |
| Octanal | 24 | 4.87 | 0.98 | 22.86 | 2.06 | 3 | |
| Nonanal | 15 | 10.86 | 0 | 34.30 | 0 | 2 | |
| Decanal | 20 | 17.00 | 0 | 78.05 | 0 | 3 | |
| Benzaldehyde | 24 | 2.26 | 0.40 | 19.22 | 1.01 | 1 | |

Table 1 Occurrence and relative abundance of volatile compounds found in 24 genotypes of Actinidia chinensis (Continued)

| Compound | Occurrence | Mean | Minimum | Maximum) /(ng g ⁻¹ FW) | 'Hort16A' | Cluster no. |
|--------------------------|-------------|----------|---------|------------------------------------|-----------|-------------|
| | (genotypes) | | | | | |
| Ethylbenzaldehyde | 24 | 14.36 | 1.86 | 180.01 | 10.99 | 4 |
| 4-Ethylbenzaldehyde | 22 | 8.14 | 0 | 155.57 | 2.60 | 4 |
| Ester | | | | | | |
| Ethyl acetate | 23 | 12.26 | 0 | 55.60 | 2.56 | 4 |
| Ethyl propanoate | 15 | 7.67 | 0 | 51.87 | 0.39 | 4 |
| Ethyl 2-methylpropanoate | 17 | 1.16 | 0 | 8.51 | 0.46 | 2 |
| Methyl butanoate | 14 | 2.46 | 0 | 26.49 | 1.33 | 4 |
| Ethyl butanoate | 21 | 75.95 | 0 | 977.65 | 42.82 | 4 |
| Butyl acetate | 1 | 0.03 | 0 | 0.73 | 0 | 1 |
| Ethyl hexanoate | 10 | 1.41 | 0 | 11.53 | 0.48 | 4 |
| Methyl benzoate | 19 | 0.68 | 0 | 1.86 | 1.33 | 3 |
| Ethyl benzoate | 21 | 1.91 | 0 | 3.97 | 3.75 | 2 |
| Methyl salicylate | 13 | 0.89 | 0 | 7.88 | 0 | 1 |
| Гегрепе | | | | | | |
| α-Pinene | 24 | 13.92 | 0.26 | 57.08 | 13.80 | 4 |
| Camphene | 7 | 0.27 | 0 | 2.41 | 0 | 4 |
| β-Pinene | 24 | 21.03 | 0.25 | 99.9 | 18.56 | 4 |
| Sabinene | 19 | 2.47 | 0 | 13.07 | 1.42 | 2 |
| α-Terpinene | 5 | 0.63 | 0 | 5.09 | 0 | 1 |
| Limonene | 16 | 1.70 | 0 | 6.06 | 0 | 2 |
| Cineole, 1,8- | 5 | 0.53 | 0 | 10.76 | 0 | 2 |
| γ-terpinene | 17 | 1.64 | 0 | 6.05 | 1.38 | 1 |
| ρ-Cymene | 20 | 2.82 | 0 | 17.35 | 2.08 | 3 |
| Terpinolene, iso- | 2 | 0.50 | 0 | 6.62 | 0 | 1 |
| Geranyl acetone | 23 | 8.88 | 0 | 113.96 | 4.19 | 3 |
| Hydrocarbon | | | | | | |
| Benzene | 24 | 5.45 | 1.36 | 13.97 | 4.76 | 1 |
| Styrene | 24 | 5.68 | 0.41 | 24.81 | 7.57 | 4 |
| Toluene | 24 | 9.29 | 4.52 | 18.92 | 12.23 | 1 |
| Trimethyl benzene | 23 | 2.27 | 0 | 5.18 | 1.47 | 2 |
| Pentandodecane | 22 | 3.64 | 0 | 10.14 | 0 | 3 |
| Alcohol | | | | | | |
| Methanol | 24 | 11.36 | 0.85 | 84.22 | 45.71 | 2 |
| Ethanol | 24 | 1 536.83 | 48.35 | 4 468.81 | 2 066. 28 | 4 |
| Propanol | 19 | 13.91 | 0 | 256.96 | 11.43 | 4 |
| 1-Methoxy-propan-2-ol | 2 | 0.25 | 0 | 4.26 | 0 | 2 |
| Butanol | 19 | 2.49 | 0 | 8.41 | 7.15 | 3 |
| Penten-3-ol | 21 | 2.602 | 0 | 12.17 | 0.69 | 3 |
| Pentanol | 23 | 2.19 | 0 | 8.03 | 0.29 | 3 |

Table 1 Occurrence and relative abundance of volatile compounds found in 24 genotypes of Actinidia chinensis (Continued)

| Compound | Occurrence | Mean | Minimum | Maximum | 'Hort16A' | Cluster no |
|-----------------------------------|-------------|---------------------------|------------------|---------------------------|--------------------------|------------|
| • | (genotypes) | /(ng g ⁻¹ FW) | /(ng g - 1 FW) | /(ng g ⁻¹ FW) | /(ng g ⁻¹ FW) | |
| Hexanol | 24 | 8.81 | 0.31 | 38.09 | 3.25 | 4 |
| Hex-Z3-enol | 18 | 12.84 | 0 | 211.41 | 7.42 | 3 |
| Hex-E2-enol | 24 | 19.71 | 1.81 | 68.42 | 6.57 | 1 |
| Ether | | | | | | |
| Tetrahydrofuran | 23 | 2.31 | 0 | 29.60 | 1.04 | 1 |
| Acid | | | | | | |
| Acetic acid | 24 | 20.74 | 5.42 | 56.97 | 12.72 | 3 |
| Ketone | | | | | | |
| Acetone | 24 | 10.62 | 4.81 | 21.17 | 9.22 | 3 |
| 2-Butanone | 23 | 2.02 | 0 | 11.50 | 0.47 | 3 |
| 3-Methylbutan-2-one | 4 | 0.74 | 0 | 15.14 | 0 | 1 |
| Butane-2,3-dione | 1 | 0.01 | 0 | 0.20 | 0 | 3 |
| Hexan-3-one | 1 | 0.05 | 0 | 1.12 | 0 | 4 |
| 2-Methylpentan-3-one ^a | 1 | 0.01 | 0 | 0.18 | 0 | 1 |
| 4-Methyl-2-hexanone ^a | 1 | 0.17 | 0 | 4.00 | 0 | 1 |
| 4-Methylpentan-2-one ^a | 1 | 0.08 | 0 | 1.96 | 0 | 1 |
| Hexen-3-one | 3 | 0.13 | 0 | 1.69 | 0 | 1 |
| 5-methylhexan-2-one ^a | 1 | 0.23 | 0 | 5.41 | 0 | 1 |
| 3-hydroxybutan-2-one | 1 | 0.013 | 0 | 0.31 | 0 | 3 |
| 6-methylhept-5-en-2-one | 23 | 1.25 | 0 | 4.28 | 0 | 3 |
| 4-hydroxy-4-methylpentan-2-one | 9 | 1.26 | 0 | 7.01 | 2.24 | 3 |
| Unknowns | | | | | | |
| Unknown 3826 (1102) | 11 | 2.51 | 0 | 15.26 | 0 | 2 |
| Unknown 160 | 11 | 1.32 | 0 | 9.17 | 0 | 3 |
| Unknown 3819 (1511) | 5 | 0.34 | 0 | 2.99 | 0 | 2 |
| Unknown 3779 (1588) | 12 | 3.22 | 0 | 71.13 | 0 | 3 |

avolatile not included in PCA

Table 2 Standardized coefficients and percentage of variation of Principal Components (PCs) for four clusters of *Actinidia chinensis* volatiles

| Volatile | C1_PC1 | C1_PC2 | C1_PC3 |
|---------------------|--------|--------|--------|
| a. Cluster 1 | | | |
| Propanal | 0.648 | -0.114 | -0.231 |
| Tetrahydrofuran | 0.944 | -0.039 | 0.074 |
| 3-Methylbutan-2-one | 0.949 | 0.027 | 0.049 |
| Benzene | 0.458 | -0.511 | -0.171 |
| 2-Methylbutanal | 0.413 | 0.574 | -0.604 |
| Toluene | 0.614 | -0.211 | -0.167 |
| Hexen-3-one | 0.904 | -0.233 | 0.010 |

Table 2 Standardized coefficients and percentage of variation of Principal

Components (PCs) for four clusters of Actinidia chinensis volatiles (Continued)

| Volatile | C1_PC1 | C1 | _PC2 | C1_PC3 |
|-------------------------------------|--------|--------|----------|--------|
| Butyl acetate | 0.948 | _ | 0.011 | 0.079 |
| α-Terpinene | 0.667 | | 0.045 | 0.439 |
| Heptanal | 0.725 | _ | 0.080 | 0.234 |
| γ-Terpinene | 0.500 | | 0.186 | 0.601 |
| Terpinolene, iso- | 0.675 | | 0.501 | -0.448 |
| Hex-E2-enol | 0.217 | | 0.819 | 0.075 |
| Benzaldehyde | 0.976 | - | 0.083 | -0.011 |
| Methyl salicylate | -0.154 | | 0.448 | 0.563 |
| Percentage of total variation/ $\%$ | 49.22 | 1 | 2.51 | 10.82 |
| Volatile | C2_PC1 | C2_PC2 | C2_PC3 | C2_PC4 |
| b. Cluster 2 | | | | |
| Butanal | -0.435 | 0.700 | -0.198 | 0.271 |
| Methanol | 0.543 | 0.251 | 0.709 | -0.051 |
| Ethyl 2-methylpropanoate | 0.336 | 0.544 | -0.592 | -0.002 |
| Unknown 3826 (1102) | 0.705 | 0.531 | 0.168 | 0.136 |
| Sabinene | -0.467 | 0.263 | 0.100 | -0.359 |
| 1-Methoxy-propan-2-ol | 0.412 | 0.325 | 0.781 | 0.031 |
| Unknown 3819 (1511) | 0.642 | 0.498 | -0.370 | -0.260 |
| Limonene | -0.672 | 0.527 | 0.039 | 0.289 |
| Cineole, 1,8- | -0.501 | 0.566 | -0.010 | -0.051 |
| Hex-E2-enal | 0.277 | 0.054 | -0.009 | 0.790 |
| Trimethyl benzene | -0.670 | 0.388 | 0.308 | -0.115 |
| Nonanal | 0.677 | 0.302 | -0.160 | -0.429 |
| Ethyl benzoate | 0.563 | -0.074 | -0.160 | 0.354 |
| Percentage of total variation/1/0 | 29.98 | 18.47 | 14.04 | 10.19 |
| c. Cluster 3 | | | | |
| Acetone | 0.739 | 0.016 | -0.103 | -0.354 |
| 2-Butanone | 0.107 | -0.409 | -0.263 | -0.417 |
| Butane-2,3-dione | -0.188 | 0.252 | 0.818 | -0.033 |
| Unknown 160 | -0.313 | 0.578 | -0.446 | 0.223 |
| Hex-Z3-enal | 0.897 | 0.326 | 0.062 | 0.071 |
| Butanol | -0.370 | 0.627 | 0.107 | -0.254 |
| Penten-3-ol | 0.743 | 0.344 | 0.097 | 0.055 |
| Pentanol | 0.619 | -0.018 | -0.001 | 0.409 |
| p-Cymene | -0.066 | 0.306 | 0.828 | -0.035 |
| Octanal | 0.949 | 0.065 | 0.093 | -0.077 |
| 3-Hydroxybutan-2-one- | 0.081 | -0.118 | -0.062 | 0.830 |
| 2-Methylbutanal | 0.413 | 0.574 | -0.604 | |
| 6-Methylhept-5-en-2-one | 0.877 | 0.038 | - 0.054 | 0.092 |
| 4-Hydroxy-4-methylpentan-2-one | -0.347 | 0.743 | - 0.288 | -0.132 |
| Hex-Z3-enol | 0.858 | 0.409 | 0.068 | 0.056 |
| Acetic acid | 0.732 | 0.007 | - 0. 281 | -0.273 |

Table 2 Standardized coefficients and percentage of variation of Principal Components (PCs) for four clusters of *Actinidia chinensis* volatiles

(Continued)

| Components (| cs) for four clusters of Actinidia chinensis volatiles | | | (Continued) | |
|---------------------------------|--|----------|--------|-------------|--|
| Volatile | C3_PC1 | C3_PC2 | C3_PC3 | C3_PC4 | |
| Pentandodecane | 0.447 | -0.213 | -0.135 | 0.201 | |
| Decanal | 0.871 | - 0. 143 | 0.031 | 0.004 | |
| Unknown 3779 (1588) | 0.894 | 0.328 | 0.063 | 0.066 | |
| Methyl benzoate | 0.601 | -0.343 | -0.006 | -0.116 | |
| Geranyl acetone | -0.144 | 0.655 | -0.455 | 0.013 | |
| Percentage of total variation/% | 38.88 | 13.68 | 10.35 | 7.24 | |
| Volatile | C4_PC1 | C4 | _PC2 | C4_PC3 | |
| d. Cluster 4 | | | | | |
| Acetaldehyde | 0.211 | - (| 0. 238 | -0.152 | |
| Ethyl acetate | 0.716 | - (|). 125 | 0.519 | |
| Ethanol | 0.628 | - (| 0.308 | 0.377 | |
| Ethyl propanoate | 0.641 | - (| 0.023 | 0.421 | |
| Methyl butanoate | 0.895 | - (| 0.012 | 0.279 | |
| Hexan-3-one | -0.098 | - (|). 113 | 0.218 | |
| α-Pinene | 0.782 | -0.012 | | -0.524 | |
| Ethyl butanoate | 0.844 | -0.031 | | 0.398 | |
| Propanol | -0.164 | 0.918 | | 0.175 | |
| Camphene | 0.809 | 0.104 | | -0.390 | |
| Hexanal | 0.583 | 0.284 | | -0.455 | |
| β-Pinene | 0.838 | (| 0.032 | -0.453 | |
| Hex-E3-enal | 0.860 | (|). 177 | 0.093 | |
| Hex-Z2-enal | 0.505 | 0.651 | | -0.331 | |
| Ethyl hexanoate | 0.652 | -0.069 | | 0.494 | |
| Styrene | 0.412 | -0.347 | | -0.189 | |
| Hexanol | 0.918 | 0.262 | | -0.109 | |
| Ethylbenzaldehyde | -0.150 | 0.907 | | 0.238 | |
| 4-Ethylbenzaldehyde | -0.154 | (|). 921 | 0.209 | |
| Percentage of total variation/% | 40.51 | 18.09 | | 11.97 | |

3 Results and discussion

3.1 Chemical analysis

The composition of the volatile compounds from ripe fruit of the 24 genotypes of *A. chinensis* is summarised in Table 1. Seventy-two volatiles (16 aldehydes, 14 ketones, 11 terpenes, 10 esters, 10 alcohols, 5 hydrocarbons, 1 ether, 1 acid and 4 unknowns) were detected and quantified. Seventeen volatiles were found in all genotypes (Table1). Of these, ten were also found in *A. arguta* (Matich et al., 2003) and four in 'Hayward' (Young et al., 1995). Most volatiles of 'Hort16A' were detected in our genotypes, except for methyl carbonate, ethyl pentanoate and ethyl octanoate, which were found at low levels in 'Hort16A' (Wang et al., 2010). Our results confirmed that, as in 'Hayward' (Paterson et al., 1991), ethanol and hex-*E*2-enal were abundant volatiles in mature fruit of *A. chinensis*. Hexanal and ethyl butanoate were other major components in these *A. chinensis* fruit. Ten of eleven major volatile flavour compounds detected in 'Hayward' (*A. deliciosa*) by Young et al.

(1995) were also detected in our A. chinensis genotypes. Methyl hexanoate was not found but ethyl hexanoate, which was not a major compound identified by Young et al. (1995), was found in ten genotypes in this study. Our results also confirmed that 'Hort16A' had a trace level of ethyl hexanoate (Wang et al., 2010).

Twelve uncommon volatiles were detected in between one and three genotypes (Table 3) and of these, seven (four ketones and one each of aldehyde, ester and terpene) were found only in genotype P2. The ester butyl acetate, found in P2, occurs in many fruits and is a constituent of apple odours (Surburg and Panten, 2006).

3.2 Multivariate analysis of volatiles

3.2.1 'Flavour impact' volatiles of A. chinensis fruit

Applying the parsimony criterion of 70 % for PCA meant it was necessary to consider only 14 PCs (Table 2), the first three or four from each of the four clusters (C1 to C4) grouped using VARCLUS based on a phenotypic correlation matrix between volatiles. Therefore, PCA offered a method to reduce the complexity of the multidimensional system, by maximizing the component loadings variance and eliminating less influential components. Moreover, the standardised coefficients of PCA revealed significant patterns in the volatile data(Table 2). In chemometric data analysis, PCA is usually applied as a first step in the analysis and the PCs often serve as the input data for other approaches, such as multivariate regression and discriminant analysis (Daszykowski et al., 2007).

In this study, PCA in combination with MLR made it possible to estimate correlations between volatiles and sensory or fruit data. MLR can establish the relative predictive importance of the independent variables by comparing standardised coefficients. Table 4 shows standardised coefficients of MLR for DM, sugars, acids and sensory data conducted as functions of the 14 PCs. Only two PCs (C2_PC3 and C4_PC1) were positively correlated with 'Hort16A'-like odour(0.456 and 0.482) and 'Hort16A'-like flavour(0.224 and 0.235). They also correlated positively with sweet taste and negatively with sour taste. C4_PC1 was correlated with ethyl butanoate flavour and odour(0.524 and 0.465), whereas C2_PC3 was correlated with ethyl butanoate odour alone(0.307). These results suggest that both PCs were associated with 'flavour impact volatiles'.

Table 3 Twelve uncommon volatiles found in genotypes of Actinidia chinensis (ng g⁻¹ FW)

| | P2 | Р3 | P4 | Р8 | P9 | P11 | P13 | P21 | P22 |
|-----------------------|------|------|------|------|------|------|------|------|------|
| 2-Methylbutanal | 1.05 | 2.54 | 0 | 0 | 0 | 0 | 0 | 0 | 1.51 |
| Butane-2, 3-dione | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0.20 | 0 |
| Hexan-3-one | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1.12 |
| 2-Methylpentan-3-one | 0.18 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 4-Methyl-2-hexanone | 4.00 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 4-Methylpentan-2-one | 1.96 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Hexen-3-one | 1.69 | 0 | 0.43 | 0.93 | 0 | 0 | 0 | 0 | 0 |
| Butyl acetate | 0.73 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 5-Methylhexan-2-one | 5.41 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 1-Methoxy-propan-2-ol | 0 | 0 | 0 | 0 | 0 | 4.26 | 1.69 | 0 | 0 |
| Terpinolene, iso- | 5.29 | 6.62 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 3-Hydroxybutan-2-one | 0 | 0 | 0 | 0 | 0.31 | 0 | 0 | 0 | 0 |

Table 4 Standardised coefficients for multiple linear regression (MLR) of Actinidia chinensis fruit and sensory attributes as a function of the 14 Principal Components (PCs)

| Components | DM | Fructose | Glucose | Sucrose | myo- Inositol | Malic acid | Quinic acid | Citric | TA | Sweet | Sour | 'Hort16A'-'Hort16A' like flavour like odour | 'Hort16A'- like odour | Ethyl butanoate flavour | Ethyl butanoate odour |
|------------|--------|----------|---------|---------|------------------|---------------|----------------|--------|--------|--------|--------|--|--------------------------|-------------------------------|-----------------------------|
| C1_PC1 | -0.174 | -0.193 | -0.225 | -0.046 | 0.104 | 0.235 | -0.120 | -0.208 | 0.025 | -0.089 | 0.205 | -0.334 | -0.289 | 0.169 | -0.106 |
| C1_PC2 | -0.074 | -0.920 | -0.766 | 0.036 | -0.135 | -0.736 | -0.600 | -0.994 | 0.440 | -0.058 | 0.285 | -0.176 | -0.203 | 0.153 | -0.249 |
| C1_PC3 | 0.062 | -0.129 | -0.180 | 0.228 | -0.165 | es I | 1 | -0.513 | -0.410 | -0.326 | 0.174 | ı | I | 0.287 | -0.559 |
| C2_PC1 | -0.081 | -0.041 | -0.277 | 0.257 | -0.049 | 0.103 | -0.252 | -0.690 | -0.251 | -0.182 | 0.263 | -0.141 | -0.425 | 0.255 | 0.018 |
| C2_PC2 | 0.184 | 0.281 | 0.262 | 0.028 | 0.321 | 0.143 | 0.628 | -0.015 | -0.230 | 0.278 | -0.307 | 0.065 | -0.035 | -0.328 | 0.427 |
| C2_PC3 | -0.261 | -0.029 | 0.057 | -0.136 | -0.140 | -0.303 | -0.204 | 0.248 | 0.004 | 0.385 | -0.311 | 0.224 | 0.456 | -0.014 | 0.307 |
| C2_PC4 | -0.249 | -0.226 | -0.293 | -0.394 | 0.637 | 0.204 | 0.048 | 0.236 | 0.483 | -0.684 | 0.680 | -0.192 | -0.427 | -0.421 | 0.299 |
| C3_PC1 | -0.284 | -0.889 | -0.831 | -0.137 | -0.456 | -0.371 | -0.795 | -0.420 | 0.591 | -0.601 | 0.802 | -0.416 | -0.216 | ı | -0.908 |
| C3_PC2 | 0.096 | -0.048 | -0.143 | 0.233 | 0.115 | -0.364 | -0.058 | -0.619 | 0.128 | 0.058 | -0.059 | 0.011 | -0.484 | -0.077 | -0.212 |
| C3_PC3 | 0.127 | 0.254 | 0.170 | -0.247 | 0.067 | 0.419 | -0.193 | 0.270 | 0.162 | -0.157 | 0.008 | 0.094 | -0.225 | -0.272 | 0.072 |
| C3_PC4 | 0.006 | 0.114 | 0.110 | 0.001 | -0.229 | 0.052 | 0.130 | 0.005 | -0.179 | -0.296 | 0.259 | -0.402 | -0.398 | -0.307 | -0.056 |
| C4_PC1 | 0.630 | 0.962 | 0.805 | 0.182 | -0.329 | 0.657 | 0.323 | 0.640 | -0.350 | 0.400 | -0.666 | 0.235 | 0.482 | 0.524 | 0.465 |
| C4_PC2 | -0.442 | -0.175 | -0.251 | -0.276 | -0.187 | -0.034 | 0.026 | 0.061 | -0.201 | -0.067 | -0.115 | 0.019 | 0.336 | 0.188 | -0.065 |
| C4_PC3 | -0.267 | -0.613 | -0.586 | -0.386 | 0.663 | -0.256 | -0.044 | -0.464 | 0.521 | -0.371 | 0.590 | -0.174 | -0.112 | -0.118 | 0.660 |

TA Titratable acidity

^a The component did not fit the regression model, e. g. caused some standardised coefficients (absolute values) to be > 1

Table 5 Standardized coefficients, percentage of variation of two Principal Components (PCs) and odour threshold in water for the 13 key *Actinidia chinensis* volatiles

| Volatile No | K_PC1 | K_PC2 | Odour threshold ppb in water |
|---------------------------------|-------|--------|------------------------------|
| Ethyl acetate | 0.109 | -0.205 | 8 000 a |
| Ethanol | 0.096 | -0.211 | 62 450 ^a |
| Ethyl propanoate | 0.096 | -0.144 | 15 ª |
| Methyl butanoate | 0.134 | -0.108 | 31 ^a |
| x-Pinene | 0.113 | 0.206 | 120° |
| Ethyl butanoate | 0.127 | -0.156 | 4.5° |
| Camphene | 0.118 | 0.185 | - |
| Hexanal | 0.082 | 0.262 | 35 ^a |
| 3-Pinene | 0.122 | 0.189 | 140^{b} |
| Hex-E3-enal | 0.127 | 0.011 | - |
| Hex-Z2-enal | 0.074 | 0.268 | - |
| Ethyl hexanoate | 0.099 | -0.193 | 1.7ª |
| Hexanol | 0.061 | -0.022 | 1 052 a |
| Percentage of total variation/% | 52.0 | 18.0 | |

^aRychlik et al., 1998; ^bFazzalari, 1978.

Thirteen volatiles in Cluster 4 and two volatiles (methanol and 1-methoxy-propan-2-ol) in Cluster 2 had moderate to high standardised PCA coefficients for C4_PC1 and C2_PC3, respectively (Table 2). C4_PC1 accounted for 40.5 % of the total variance in Cluster 4, whereas C2_PC3 accounted for only 14.0 % of the total variance in Cluster 2. The first component, as usual in PCA, was the most important, accounting for most of the total variance (Morrison, 1990). The results imply that the potential 'flavour impact volatiles' in Cluster 4 might be more important contributors to 'Hort16A'-like flavour and odour than those in Cluster 2. Methanol and 1-methoxy-propan-2-ol in Cluster 2 are alcohols and 1-methoxy-propan-2-ol was detected in only two genotypes (Table 1). Free and esterified, saturated primary alcohols occur widely in fruits, but their odour is relatively weak (Surburg and Panten, 2006). However, these alcohols could be involved in AAT activities to produce esters (Ke et al., 1994).

The 13 volatiles in Cluster 4 identified as potential 'flavour impact' volatiles for A. chinensis fruit included five esters (ethyl acetate, ethyl propanoate, methyl butanoate, ethyl butanoate and ethyl hexanoate), three terpenes (α-pinene, camphene and β-pinene), three aldehydes (hexanal, hex-E3-enal and hex-Z2-enal) and two alcohols (ethanol and hexanol) (Tables 1,2). These volatiles together represented 66.0% of the total volatiles produced by these A. chinensis fruit. For some genotypes, the concentrations of ethyl propanoate, ethyl butanoate, ethyl hexanoate and hexanal were higher than their odour thresholds (Table 5) (Rychlik et al., 1998). A GC-sniffing study showed that acetaldehyde, hexanal, ethyl butanoate and hex-E2-enal had odour activity in macerated fruit of 'Hort16A' (Friel et al., 2007). Young et al. (1983) reported that hex-E2-enal, ethyl butanoate, methyl benzoate and hexanal were important contributors to the aroma and flavour of 'Hayward' (A.

deliciosa). Studies on the causal effects of selected flavour volatiles on the intensity of sensory attributes in 'Hayward' showed that ethyl butanoate, methyl benzoate and hexanal had positive correlations with sweet aroma and flavour (McMath et al., 1991; Young et al., 1995). The most pronounced effect on 'kiwifruit aroma and flavour' was produced by ethyl butanoate (Gilbert et al., 1996). Paterson et al. (1991) reported that ethyl butanoate, the major ester present in 'Hayward', appeared to be a predictor of flavour acceptability. Our results also showed that ethyl butanoate was the major ester present in these A. chinensis fruit. However, high concentrations of this volatile can also contribute to a 'very ripe kiwifruit-like' perception of flavour (Paterson et al., 1991; Gilbert et al., 1996), which is not a desirable attribute (Stec et al., 1989). Thus, the balance of this volatile with others may be a more important characteristic. An integrated approach using GC-MS and GC-O is needed to confirm whether these 13 volatiles play a significant role in contributing to the flavour and odour of A. chinensis fruit.

C4_PC2 was also correlated with 'Hort16A'-like odour, but not with 'Hort16A'-like flavour, ethyl butanoate flavour or odour, or sweet or sour tastes (Table 4). Propanol, ethylbenzaldehyde and 4-ethylbenzaldehyde were highly correlated with C4_PC2 (Table 2). This indicated that C4_PC2 identified 'Hort16A'-like odour volatiles'. Turin(1996) found that ethylbenzaldehyde had a typical bitter almond odour and Pennarun et al. (2003) found that 4-ethylbenzaldehyde was characterized by a minty and aniseed odour. Propanol is an alcohol with relatively weak odour and its concentration (Table 1) was below its odour threshold (Rychlik et al.,1998). As C4_PC2 accounted for only 18.1 % of the total variance in Cluster 4, it was not an important component compared with C4_PC1 (40.5 %). However, volatiles represented in this cluster (C4_PC2) could affect the overall perception of flavour or odour by altering the complexity of the experience. Previous studies reported that adding sub-and perithreshold odourant can have a measurable impact on perceived flavour or aroma (Atanasova et al., 2005; Labbe et al., 2007; Miyazawa et al., 2008).

Ethyl 2-methylpropanoate and unknown 3819 in Cluster 2 had negative correlations with C2_ PC3. Thus, genotypes with a high concentration of these volatiles had less 'Hort16A'-like flavour and odour, and 'Hort16A' (P24) had a low concentration of these volatiles. Ethyl 2-methylpropanoate was present in 17 genotypes, whereas unknown 3819 (1511) was found in only five genotypes (Table 1). The concentrations of ethyl 2-methylpropanoate ranged from 0.23 to 8.51 ng g⁻¹ FW, higher than the odour threshold of 0.1 µg 1⁻¹ in water (Takeoka et al., 1990). Matich et al. (2003) reported that the aroma of fruit of selection A3 (A. arguta) was probably the most intense because of the very sweet, fruity odours of ethyl 2-methylpropanoate and ethyl 2-methylbutanoate. Frank et al. (2007), using GC/MS-O, reported that ethyl-2-methylpropanoate ('melon, bubblegum') was the most potent odour-active ester compound in 'Hayward'. Ethyl 2-methylpropanoate was described as a characteristic fruity volatile in pineapple (Tokitomo et al., 2005) and blackberries (Klesk and Qian, 2003). Further study is needed to explore whether ethyl 2-methylpropanoate may contribute different and positive flavours and odours to A. chinensis fruit, and therefore be a desirable compound in a breeding population. From a study on consumers' preferences for kiwifruit, Wismer et al. (2005) suggested that fruit breeding should target not only elite fruit that are significantly more like existing successful cultivars, but also unique fruit that could create major new flavour niches.

C4_PC3 had low negative correlations with 'Hort16A'-like flavour odour, but a high positive correlation with ethyl butanoate odour and 'sour taste' (Table 4). This component also had positive moderate correlations with ethyl esters (ethyl acetate, ethyl propanoate, ethyl butanoate and ethyl hexanoate) and an alcohol (ethanol), but negative moderate correlations with terpenes (α-pinene, camphene and β-pinene) and two aldehydes (hexanal and hex-Z2-enal) (Table 2). All volatiles correlated with C4_PC3 were found to be potential 'flavour impact volatiles' as in C4_PC1 (as discussed above). Ethyl butanoate was found in much higher concentrations in ripe fruit than in firm fruit of 'Hort16A', and this was shown to be a feature of ripening stage rather than physical maceration of tissue, and hence either enzyme activity or substrate availability (Friel et al., 2007). Marsh et al. (2003) found that a rise in off-flavours and acid taste was consistent with the addition of malate to 'Hayward' pulps. It was suggested that the release of alcohols was associated with a change in acidity (Marsh et al., 2006). In a study of 'Hayward' fruit in an ethylene-free environment, Burdon et al. (2005) suggested that the production of significant esters was driven by alcohol metabolism that developed as fruit reached and passed through the eating firmness range; and Paterson et al. (1991) demonstrated an association between ester concentrations and increasing fruit ethylene production in 'Hayward' fruit. The results indicated that the higher concentrations of esters could relate to over-ripe fruit with strong ethyl butanoate odour and 'sour taste'. Thus, genotypes with overripe fruit would be expected to have a high positive score on C4_PC3. The results suggest that C4_PC3 identified 'kiwifruit fruit ripeness', which is probably a measure of the balance between different components of flavours and odours. The presence of this component may also suggest that AAT enzymes with ethanol may play a key role in the biosynthesis of ethyl esters (Ke et al., 1994), and that alcohol dehydrogenase (ADH) and lipoxygenase (LOX) enzymes could play specific roles in the regulation of odour biosynthesis (Zhang et al., 2009; González-Agüero et al., 2009). Sequence analysis of acyltransferases (ATs) from the Actinidia EST database showed that two clades contained enzymes involved in the synthesis of flavour-related esters (Crowhurst et al., 2008). Further study is needed to explore biosynthetic pathways leading to volatile production.

In this study, two PCs(C2_PC4 and C3_PC1) had high correlations with 'sour taste' positively and 'sweet taste' negatively(Table 4). C3_PC1 also had a very high negative correlation with ethyl butanoate odour. This group of volatiles included hex-E2-enal in Cluster 2 and hex-Z3-enal, octanal, 6-methylhept-5-en-2-one, hex-Z3-enol, decanal and the unknown 3779 (1588) in Cluster 3. The mean concentrations of hex-E2-enal, hex-Z3-enal, octanal and decanal (Table 1) were higher than their odour thresholds (Rychlik et al., 1998). Hex-E2-enal was the major volatile component in mature fruit of 'Hayward' (Bartley and Schwede, 1989). Gilbert et al. (1996) studied the consumer perception and acceptability of three volatiles at variable levels in a model base solution. They found that hex-E2-enal had a negative effect on the acceptability attributes (i. e., 'overall liking', 'liking of aroma' and 'liking of flavour'), but a positive correlation with the perceived intensity of 'kiwifruit aroma' and 'acid flavour'. Frank et al. (2007) reported that hex-E2-enal (perceived as marzipan, sweet) was the odour compound with the greatest impact in 'Hayward'. In apricot, hex-E2-enal gave a grassy note, whereas 6-methylhept-5-en-2-one developed a floral note (Guillot et al., 2006). Friel et al. (2007) reported that a low concentration of 6-methylhept-5-en-2-one was found in

the firm fruit of 'Hort16A'. It is well known that the volatiles and sensory properties of kiwifruit are markedly affected by fruit ripeness (Stec et al., 1989). Thus, this group of volatiles could be related to a predominance of aldehydes and associated with less ripe fruit.

C2_PC1 and C2_PC4 had moderate negative correlations with 'Hort16A'-like odour, while C2_PC2 and C2_PC4 had a moderate positive correlation with ethyl butanoate odour but a moderate negative correlation with ethyl butanoate flavour (Table 4). Most PCs for Clusters 1 and 3 had negative correlations with 'Hort16A'-like flavour and odour, and ethyl butanoate flavour and odour, except for C3_PC2 and C3_PC3, which had very low positive correlations with 'Hort16A'-like flavour (Table 4). C3_PC3 also had a very low positive correlation with ethyl butanoate odour. All volatiles in Clusters 1 and 3 had moderate to high correlations with PCs (Table 2). The results showed that most volatiles in Cluster 2 (except for methanol, ethyl 2-methylpropanoate and 1-methoxy-propan-2-ol) were correlated with 'atypical 'Hort16A'-like odour' and those in Clusters 1 and 3 were associated with 'atypical kiwifruit flavour' in A. chinensis fruit.

Most of the aldehyde compounds were in Clusters 1 ~ 3 (Table 1), although some were grouped into Cluster 4 as flavour volatiles (hexanal, hex-E3-enal and hex-Z2-enal) or odour volatiles (ethylbenzaldehyde and 4-ethylbenzaldehyde), as discussed above. The lower fatty aldehydes C₂ ~ C₇ impart fruity characters to flavour compositions (Surburg and Panten, 2006). The same authors also reported that octanal had a pungent odour that became citrus-like on dilution, nonanal had a fatty rose-like odour and decanal a strong odour reminiscent of orange peel. While these aldehydes had negative correlations with 'Hort16A'-like flavour and odour, they could contribute to breeding interesting new flavours in A. chinensis because their concentrations were higher than their odour thresholds (Rychlik et al., 1998). However, Friel et al. (2007) pointed out that as the volatiles in kiwifruit studies were generally collected over a period of 20 min (15 min for this study), it is unclear whether concentrations of volatile lipid oxidation products (e. g., aldehydes) would contribute significantly to the odour experienced by a consumer while eating a piece of fruit (i. e., over 5 ~ 15 s).

All ketone compounds were in Clusters 1 and 3, except for hexan-3-one in Cluster 4 (Table 1). However, hexan-3-one had low standardized coefficients for the three C4_PCs (Table 2), suggesting that it might not be an important flavour compound in *A. chinensis* fruit. Moreover, only three ketones (acetone, 2-butanone and 6-methylhept-5-en-2-one) were common in *A. chinensis*, being detected in 23 or 24 genotypes. The concentrations of acetone and 6-methylhept-5-en-2-one were lower than their odour thresholds for all genotypes (Rychlik et al., 1998; Buttery et al., 1990). 2-Alkanones (C3 ~ C15) have been found in the volatile fractions of many fruits and foodstuffs, but they do not contribute significantly to their odour (Surburg and Panten, 2006).

In this study, 13 volatiles were successfully identified as potential key 'flavour impact' volatiles for A. chinensis fruit from the composition of 72 volatiles by using multivariate analysis, suggesting that it can be possible to measure volatiles in a large breeding population. Moreover, MLR showed that C4_PC1 was highly correlated with other fruit attributes, e. g., DM, fructose, glucose, malic and citric acids (Table 4). In breeding, it is sometimes more convenient or effective to select for a correlated character than to select for the desired character itself (Falconer and Mackay, 1996). Methods commonly used for identifying volatile compounds, such as GCMS, require expensive equipment and the analysis of large numbers of samples is time consuming. As DM was highly

convenient and efficient alternative to direct selection for increased flavour impact volatiles in this A. chinensis population. DM is quick and easy to measure on large numbers of samples and had a positive genetic correlation with the amounts of sugars (Cheng et al., 2004). A recent meta-analysis of kiwifruit sensory experiences demonstrated unequivocally that consumers prefer high DM fruit of both 'Hayward' and 'Hort16A' (Harker et al., 2009).

3.2.2 Differentiation of genotypes associated with flavour impact volatiles

To refine differentiation between genotypes, the patterns of the 13 potential key 'flavour impact' volatiles (from Cluster 4) for the 24 genotypes were analyzed by PCA. Based on the parsimony criterion, only two PCs were considered. K_PC1 accounted for 52.0% and K_PC2 for only 18.0% of the total variance (Table 5). K_PC1 was positively correlated with all 13 volatiles, suggesting that the first component was associated with 'total key volatiles'. Thus, a genotype with high concentrations of these volatiles would be expected to have high positive scores. Particularly, genotypes with high concentrations of methyl and ethyl butanoates, hex-E3-enal and the three monoterpenes (α -pinene, camphene and β -pinene) would have higher positive scores, as indicated by higher coefficients. The results of MLR of the K_PCs as a function of sensory attributes showed that K_PC1 had a high positive correlation with ethyl butanoate flavour and a low correlation with ethyl butanoate odour (Table 6). There was also a contrast between 'Hort16A'-like flavour and 'sour taste', as indicated by negative coefficients for the first PC.

K_PC2 had a negative correlation with five esters (ethyl acetate, ethyl propanoate, methyl butanoate, ethyl butanoate and ethyl hexanoate) and ethanol, while it had a positive correlation with two aldehydes (hexanal and hex-Z2-enal) and the three monoterpenes (Table 5). The results of MLR also showed that K_PC2 had a negative correlation with 'sour taste', 'Hort16A'-like odour and flavour and ethyl butanoate flavour, but a positive correlation with ethyl butanoate odour (Table 6). K_PC2 revealed two welldefined groups; one positive and one negative for the flavour impact volatiles of 'Hort16A'-like flavour and odour. Five esters were significant contributors to 'Hort16A'-like flavour and odour. Genotypes with high concentrations of positive 'flavour impact' volatiles would have low negative scores in K_PC2, whereas genotypes with high concentrations of negative 'flavour impact' volatiles would have high positive scores. The results suggest that K_PC2 identified genotypes with 'Hort16A'-like flavour and odour'. However, a lower score for K_PC1 or K_PC2 could relate to over-ripe fruit inducing high 'sour taste' (as discussed above). Thus, genotypes with medium-low scores for K_PC1 and K_PC2 may have high 'Hort16A'-like flavour and odour and yet to some extent a 'sour taste' as well. Jaeger et al. (2003) found that consumers accepted A. chinensis fruit because they had high sweet and low acidic flavour.

Table 6 Standardised coefficients for multiple linear regression(MLR) of K_PCs as a function of *Actinidia chinensis* sensory attributes

| Sensory attributes | K_PC1 | K_PC2 | Sensory attributes | K_PC1 | K_PC2 |
|-----------------------|--------|--------|-------------------------|--------|--------|
| Ethyl butanoate odour | 0.161 | 0.109 | Sour taste | -0.221 | -0.579 |
| 'Hort16A' -like odour | 0.008 | -0.272 | Ethyl butanoate flavour | 0.444 | -0.153 |
| Sweet taste | -0.053 | -0.009 | 'Hort16A'-like flavour | -0.359 | -0.259 |

In this study, PCA has provided some insights into 'kiwifruit fruit ripeness' and an indication of the balance of different volatile components that will affect consumer responses. Those genotypes with a high positive score in K_PC2 could have an imbalance in biosynthetic pathways or lack a critical step for ester formation during fruit ripening. Further study is needed to explore differences in the biosynthetic pathways of volatile production between *A. chinensis* genotypes, to establish a biochemical basis for selecting new cultivars with flavours that appeal to consumers.

Genotypes were highly differentiated by their 'flavour impact' volatiles (Fig. 1). P1, P6, P22 and female parents B and C ('Hort16A') had medium-low scores of K_PC1 and K_PC2, which may result in fruit with high 'Hort16A'-like flavour/odour and to some extent 'sour taste'. Hence, these genotypes may be useful as parents to improve 'Hort16A'-like flavour/odour. P4, P8 and P13 had high concentrations of both positive and negative 'flavour impact' volatiles, whereas P3 had high concentrations of only negative volatiles. Four genotypes (P7, P14, P16 and P20) had low concentrations of total 'impact flavour' volatiles. Other genotypes had either medium-high concentrations of negative 'flavour impact' volatiles or medium-low concentrations of positive volatiles.

In conclusion, extended use of multivariate analysis proved very powerful for extracting maximum information from the limited plant material. In this study, volatiles were successfully grouped, using hierarchical clustering of variables based on phenotypic correlations between volatiles, to avoid a singular correlation matrix of the data in PCA. Based on these groups, using PCA and MLR, correlations between volatiles, sensory and fruit attributes were explored. However, it must be noted that the presence of a correlation does not necessarily mean that a cause and effect relationship exists. In this case, the PCs reflect the influence of different categories of volatile variables of different biological significance. One of the major strengths of PCA is to allow identification of important factors leading to biological interpretations of key processes. In this study, standardized coefficients of MLR enabled the PCs to have meaningful biological interpretations. Consequently, potential key 'flavour impact' volatiles were identified, which may make a substantial contribution to the fruit flavour of these A. chinensis genotypes. Five esters were identified that may contribute significantly to 'Hort16A'-like flavour and odour, whereas likely negative 'flavour impact' volatiles included three monoterpenes and two aldehydes. Some volatiles associated with 'sour taste', 'fruit ripeness' 'atypical 'Hort16A'-like odour and 'atypical kiwifruit flavour' were also recognized. Furthermore, the PCs provided a measure of the balance of complex volatile components that is likely to affect consumer responses. The differentiation of genotypes associated with 'flavour impact' volatiles was successfully illustrated, leading to the identification of potentially superior parents for flavour breeding. In addition, the PCA provided some insights into possible biosynthetic pathways of volatile production. However, our results confirmed that indirect selection using DM is a convenient and efficient alternative to direct selection for improving flavour volatiles of fruit (Cheng et al., 2004) providing no new flavour profiles are desired. This fits very well with recent evidence that DM has a desirable and positive influence on consumer acceptance of kiwifruit (Harker et al., 2009). However, a selection strategy based on DM also needs to take into account its positive correlation with some negative 'flavour impact' volatiles. While it is recognised that consumers will vary in their ability to detect volatile concentrations (Friel et al.,2007), we showed that our *A. chinensis* gene pool contains a variety of flavour and/or odour volatiles useful for breeding kiwifruit cultivars with new flavour characteristics. Based on our results, we can target the testing of new combinations of potential flavour impact volatiles of kiwifruit using in vitro or pulp systems (Marsh et al.,2006).

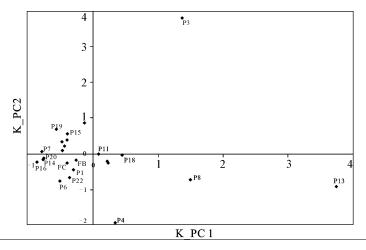


Fig. 1 Scatter plot of K_PC1 against K_PC2 for key 'flavour impact' volatiles in 24 genotypes of Actinidia chinensis

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